



## Comparison of iron deposition in subcortical structures of brains with Parkinsonian syndromes: A post-mortem 7.0-tesla magnetic resonance study

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### Abstract

It is not known whether iron (Fe) accumulation is a secondary event in the cascade of the neuronal degeneration rather than the primary cause in different Parkinsonian diseases. The purpose of the present post-mortem study is to evaluate and compare the Fe content in brains of different parkinsonian syndromes with normal control brains. Twenty-six normal brains were compared to 7 with idiopathic Parkinson's disease (PD), 18 with progressive supranuclear palsy (PSP), 18 with Lewy body dementia (LBD), 6 with corticobasal degeneration (CBD) and 5 with multiple system atrophy (MSA). The hippocampus and 11 subcortical structures were submitted to a 7.0-tesla magnetic resonance imaging (MRI). T2 and T2\* sequences were performed: the degree of T2\* hypo-intensity, representing the degree of Fe content in the different brain regions, was determined semi-quantitatively. No significant differences in Fe content were observed in all examined structures between normal brains and those with PD, PSP, LBD and CBD. In MSA there was a selective hypo-intensity on T2\* sequence in the putamen, reflecting Fe increase, with on the other hand a significant Fe decrease at the level of the dentate nucleus of the cerebellum. The present post-mortem MRI study confirms that the increased hypo-intensity in the putamen is a diagnostic hallmark of MSA, but that also the low Fe content at the level of the dentate nucleus has also to be considered as a valuable indicator of this disease.

**Keywords:** Neuropathology, 7.0-tesla MRI, brain iron deposition, Idiopathic Parkinson's disease, progressive supranuclear palsy, Lewy body dementia, corticobasal degeneration, multiple system atrophy

### 1. Introduction

In the substantia nigra the main iron (Fe) compound in dopamine and norepinephrine neurons is the neuromelanin-iron complex that serves to trap Fe and to provide neuronal protection for oxidative stress <sup>[1]</sup>.

In normal brains Fe concentrations are high in striatum, red nucleus and substantia nigra <sup>[2]</sup> but also around the dentate nucleus of the cerebellum <sup>[3]</sup>. The highest Fe concentrations in the basal ganglia are found in the globus pallidus, followed by the putamen, caudate nucleus, thalamus and white matter <sup>[4]</sup>.

Ferroptosis is a unique form of programmed cell death, characterised by cytosolic accumulation of Fe, lipid peroxides and their metabolites, affecting the plasma membrane <sup>[5]</sup>.

In patients with atypical Parkinsonian syndromes brain Fe accumulation has been extensively investigated with magnetic resonance imaging (MRI) <sup>[6]</sup>. However, "in vivo" MRI observations have to be validated by post-mortem examination <sup>[7]</sup>. T2 and T2\*- weighted MRI images are currently the preferred methods to visualize and quantify the amount of Fe in the deep brain structures <sup>[8, 9]</sup>.

The present post-mortem 7.0-tesla MRI study investigates and semi-quantifies the amount of Fe in different structures of the brains of patients with Parkinson's disease and atypical parkinsonian syndromes compared to normal controls.

### 2. Material and methods

Fifty-four patients with a Parkinsonian syndrome, who had

been followed up at the Lille University Hospital, and 26 without a neurological disease underwent an autopsy. A previously obtained informed consent from the nearest family allowed an autopsy for diagnostic and scientific purposes. The brain tissue samples were acquired from the Lille Neuro-Bank of the Lille University, federated to the "Centre des Ressources Biologiques" that acted as an institutional review board.

On post-mortem examination 7 brains were diagnosed with idiopathic Parkinson's disease (PD), 18 with progressive supranuclear palsy (PSP), 18 with Lewy body dementia (LBD), 6 with corticobasal degeneration (CBD) and 5 with multiple system atrophy (MSA).

The refined published neuropathological criteria were used for the assessment of idiopathic PD <sup>[10]</sup>. The post-mortem diagnosis of PSP was made according to the NINDS neuropathological criteria for Steele-Richardson-Olszewski syndrome (progressive supranuclear palsy) <sup>[11]</sup>. LBD was diagnosed according to the report of the consortium on DLB international workshop <sup>[12]</sup>. The diagnostic criteria of CBD were those proposed by the international consortium of behavioral neurology <sup>[13]</sup>. The neuropathological diagnosis of MSA was made according to the second consensus statement on the diagnosis of multiple system atrophy <sup>[14]</sup>.

A 7.0-tesla MRI Bruker BioSpin SA was used with an issuer-receiver cylinder coil of 72 mm inner diameter (Ettlingen, Germany), according to a previously described method <sup>[15]</sup>. Three coronal sections of a cerebral hemisphere were submitted to T2 and T2\* MRI sequences: a frontal one, a central one and one at the level of the parietal lobe.

Also a horizontal section through brainstem and a cerebellar hemisphere was examined. The severity of the Fe deposition was semi-quantitative rating on the T2\*-weighted images: no hypo-intensity (R0), small scattered hypo-intensity signals (R1), confluent zones of hypo-intensity (R2) and homogeneous dens hypo-intensity of the whole structure (R3).

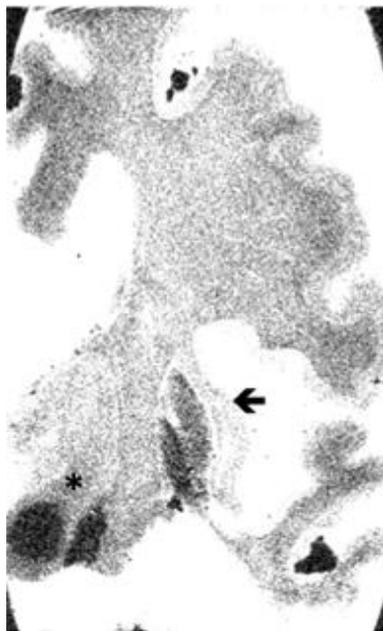
Univariate comparisons of unpaired groups were performed with the Fisher's exact test for categorical data. The non-parametric Mann-Whitney U-test was used to compare continuous variables. The significance level, two-tailed, was set at  $\leq 0.001$  for highly significant,  $\leq 0.01$  for significant and  $\leq 0.05$  for marginally significant.

**3. Results**

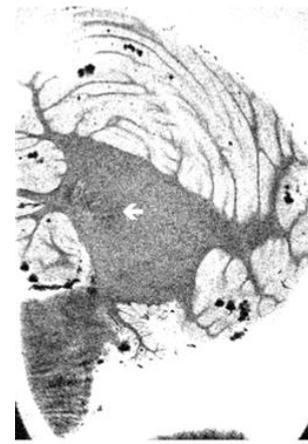
The patients with normal post-mortem brains had an average age of 66 (SD: 12) years, which was statistically not different from the 72 (SD: 5) years of the CBD and the 69 (SD: 8) years of the MSA brains. In the other groups the average age at death was older: 76 (SD: 8) years in PD ( $p \leq 0.05$ ), 79 (SD: 7) years in PSP ( $p \leq 0.01$ ) and 82 (SD:8) years in LBD ( $p \leq 0.01$ ). Gender distribution was different with a male predominance of 85% in the LBD group, compared to the normal and other parkinsonian groups, ranging from 43 until 60% ( $p \leq 0.05$ ).

Mild additional Alzheimer features were present in 12 % of PSP and in 24% of LBD brains. Moderate cerebral amyloid angiopathy was only present in 18% of the LBD group.

Semi-quantitative evaluation of the Fe content in the hippocampus and the main subcortical structures did not show any differences between the normal brains and those with PD, LBD, PSP and CBD. In the MSA brains a significant increase of Fe was only observed in the putamen ( $p \leq 0.01$ ), while a moderate decrease of Fe was observed at the level of the dentate nucleus ( $p \leq 0.05$ ) (table 1) (Fig 1-2).



**Fig 1:** T2\* MRI sequence of a coronal section of a cerebral hemisphere of a brain with multiple system atrophy. There is a selective hypo-intensity of the putamen (black arrow). The hypointensity in the globus pallidus is within normal range. Note the preserved Fe content in the substantia nigra and the red nucleus (\*).



**Fig 2:** T2\* MRI sequence of a horizontal section of a cerebellar hemisphere of a brain with multiple system atrophy. No hypo-intensity is observed at the level of the dentate nucleus (white arrow).

**Table 1:** Semi-quantitative Fe content evaluation (standard deviation) of normal brains and of brains of patients with idiopathic Parkinson disease (PD), progressive supranuclear palsy (PSP), Lewy body dementia (LBD), corticobasal degeneration (CBD) and multiple system atrophy (MSA).

Items	Normal (n = 26)	PD (n = 7)	PSP (n = 18)	LBD (n = 18)	CBD (n = 6)	MSA (n = 5)
Hippocampus	0.3(0.7)	0.0(0.0)	0.2(0.5)	0.0(0.0)	0.0(0.0)	0.0(0.0)
Clastrum	0.3(0.6)	0.0(0.0)	0.4(0.9)	0.4(0.5)	0.0(0.0)	0.0(0.0)
Caudate nucleus	0.7(0.8)	1.0(1.0)	1.4(1.0)	1.4(1.1)	0.6(0.9)	0.6(0.5)
Putamen	1.4(1.1)	1.8(0.4)	1.7(1.1)	1.9(1.1)	1.2(0.4)	3.0(0.0)**
Globus Pallidus	2.0(1.2)	1.8(0.4)	1.7(1.5)	1.9(1.3)	1.6(0.5)	3.0(0.0)
Thalamus	0.6(1.0)	0.4(0.6)	0.5(0.6)	0.4(0.5)	1.0(0.0)	0.2(0.4)
Mamillary body	0.9(1.1)	1.0(0.7)	1.6(1.2)	0.7(1.0)	0.8(0.4)	0.6(0.5)
Geniculate body	1.3(1.2)	1.0(0.7)	1.2(1.1)	1.7(1.4)	0.8(0.4)	1.2(0.8)
Subthalamic nucleus	1.0(1.3)	1.6(0.6)	1.1(1.1)	0.8(0.9)	0.8(0.5)	1.6(0.9)
Red nucleus	2.5(0.7)	2.2(0.8)	1.8(1.2)	3.0(0.0)	2.0(0.0)	3.0(0.0)
Substantia Nigra	2.6(0.5)	2.2(0.8)	2.3(1.0)	3.0(0.0)	2.0(0.0)	3.0(0.0)
Dentate nucleus	2.0(1.2)	1.4(0.9)	1.5(1.2)	1.4(1.3)	1.4(0.5)	0.6(0.5)*

**4. Discussion**

The present study only shows an increased level of Fe in the putamen and a decrease at the level of the dentate nuclei in brains with MSA. No significant differences in Fe content are observed in the dentate nuclei and the different subcortical structures between the normal and PD, PSP, LBD and CBD brains. One prejudice of this study can be due to the relative low number of patients in some of the disease groups.

Previous studies have already demonstrated Fe accumulation in the substantia nigra of PD brains, mainly in the severe forms of the disease [16]. However this does not influence the increased signal intensity on T1-weighted MRI sequence [17]. In the early stages of PD it is found to be restricted to the pars compacta of the substantia nigra [18]. In advanced stages Fe deposition extends to the pars reticularis of the substantia nigra, the red nucleus and the globus pallidus [19]. As we did not compare early and late stages of PD in this limited series, this can explain why no differences in Fe content could be observed compared to the other groups.

In LBD, similar to other studies, we did not observe an increase of Fe in the basal ganglia and in the substantia nigra [7, 20]. Only one study reported a selective Fe increase in the substantia nigra [21].

There are some controversial results concerning the Fe

accumulation in PSP brains. A post-mortem study without MRI showed a higher Fe burden in the cerebral peduncles and substantia nigra [22]. An “in vivo” MRI study showed higher R2\* values in basal ganglia, substantia nigra, and subthalamic and dentate nuclei [23]. These difference in findings can perhaps be explained by the frequent observed micro-bleeds in brainstem and cerebellum, who can be confounded with Fe deposition on T2\* MRI sequences [24].

There are only sporadic case reports concerning Fe accumulation in brains with CBD, one of them showing putaminal signal hypo-intensity on T2 weighted MRI [25]. In another neuropathological study without MRI correlates widespread Fe accumulation was observed [26].

In the parkinsonian variant of MSA (MSA-P) many MRI studies show selective Fe accumulation in the putamen, contributing to the clinical diagnosis and allowing to distinct this disease from PSP [27, 32]. The loss of Fe at the level of the dentate nucleus of the cerebellum, observed in the present study, has not previously been reported. When additional cerebellar features are present in the parkinsonian variant of MSA pontine and cerebellar atrophy is seen corresponding to the distribution of the degenerative changes. The dentate nuclei and surrounding white matter are overall more or less preserved in both the parkinsonian as in the olivo-ponto-cerebellar types of MSA [33].

Brain Fe can be selective decreased in the dentate nucleus and the thalamus of patients with idiopathic restless legs syndrome, which is considered as a brain Fe deficiency disease [34]. The only progressive cerebellar disease with absence of MRI T2\* hypointensity due to Fe loss in the dentate nucleus is the ataxia with oculomotor apraxia type 2 [35, 38]. Although this is a slowly progressing disease starting at young age it has similar clinical features as MSA. Even additional nigrostriatal involvement can be present [39].

There is no clear explanation why there is Fe loss in the dentate nucleus, while increased in the atrophic putamen of MSA brains.

## 5. Conclusion

Although the exact cause of the low Fe content in the dentate nucleus is not known it can be considered as an additional MRI diagnostic marker of MSA.

## 6. Disclosure

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## 7. References

- Zucca FA, Segura-Aguilar J, Ferrari E, Muñoz P, Paris I, Sulzer D. *et al.* Interactions between iron, dopamine and neuromelanin pathways in brain aging and Parkinson's disease. *Prog Neurobiol*. 2017; 155:96-119.
- Bilgic B, Pfefferbaum A, Rohfing T, Sullivan EV, Adalsteinsson E. MRI estimates a brain iron concentration in normal aging using quantitative susceptibility mapping. *Neuroimage*. 2012; 59:2625-2636.
- Popescu BE, Robinson CA, Rajput A, Raiput AH, Harder SL, Nichol H. *et al.* Iron, copper and zinc distribution of the cerebellum. *Cerebellum*. 2009; 8:74-79.
- Langkammer C, Krebs N, Goessler W, Scheurer E, Ebner F, Yen K *et al.* Quantitative MR imaging of brain iron: a postmortem validation study. *Radiology*. 2010; 257:455-462.
- Morris G, Berk M, Carvalho AF, Maes M, Walker AJ, Puri BK. *et al.* Why should neuroscientists worry about iron? The emerging role of ferroptosis in the pathophysiology of neurodegenerative diseases. *Behav Brain Res*. 2018; 341:154-157.
- Lee JH, Lee MS. Brain iron accumulation in atypical parkinsonian syndromes: in vivo MRI evidences for distinctive patterns. *Front Neurol*, 2019. doi: 103389/fneur. 2019.00074.
- Kitao S, Matsusue E, Fujii S, Miyoshi F, Kaminou T, Kato S. *et al.* Correlation between pathology and neuromelanin MR imaging in Parkinson's disease and dementia with Lewy bodies. *Neuroradiology*, 2013; 55:947-953.
- Pietracupa S, Martin-Bastida A, Piccini P. Iron metabolism and its detection through MRI in parkinsonian disorders: a systematic review. *Neurol Sci*. 2017; 38:2095-2101.
- Yan F, He N, Lin H, Li R. Iron deposition quantification: applications in the brain and liver. *J Magn Reson Imaging*. 2018; 48:301-317.
- Dickson DW, Braak H, Duda JE, Duyckaerts C, and Gasser T, Halliday GM. *et al.* Neuropathological assessment of Parkinson's disease: refining the diagnostic criteria. *Lancet Neurology*. 2009; 8:1150-1157.
- Hauw JJ, Daniel SE, Dickson D, Horoupian DS, Jellinger K, Lantos PL. *et al.* Preliminary NINDS neuropathological criteria for Steele-Richardson-Olszewski syndrome (progressive supranuclear palsy). *Neurology*. 1994; 44:2015-2019.
- McKeith IG, Dickson DW, Lowe J, Emre M, O'Brien JT, Feldman H. *et al.* Diagnosis and management of dementia with Lewy bodies: the third report of the LBD Consortium. *Neurology*. 2005; 65:1863-1872.
- Armstrong MJ, Litvan I, Lang AE, Bak TH, Bhatia KP, Borroni B. *et al.* Criteria for the diagnosis of corticobasal degeneration. *Neurology*. 2013; 80:496-503.
- Gilman S, Wenning GK, Low PA, Brooks DJ, Mathias CJ, Trojanowski JQ. *et al.* Second consensus statement on the diagnosis of multiple system atrophy. *Neurology*. 2008; 71:670- 676.
- De Reuck J, Auger F, Cordonnier C, Deramecourt V, Durieux N, Pasquier F. *et al.* Comparison of 7.0-T T2\* magnetic resonance imaging of cerebral bleeds in post-mortem brain sections of Alzheimer patients with their neuropathological correlates. *Cerebrovasc Dis*. 2011; 31:511-517.
- Riederer P, Sofic E, Rausch WD, Schmidt B, Reynolds GP, Jellinger K. *et al.* Transition methals, ferritin, glutathione and ascorbic acid in parkinsonian brains. *J Neurochem*. 1998; 52:515-520.
- Kitao S, Matsusue E, Fujii S, Miyoshi F, Kaminou T, Kato S. *et al.* Correlation between pathology and neuromelanin MR imaging in Parkinson's disease and dementia with Lewy bodies. *Neuroradiology*. 2013; 55:947-953.
- Jellinger K, Paulus W, Grundke-Iqbal I, Riederer P, Youdim MB. *et al.* Brain iron and ferritin in

- Parkinson's and Alzheimer's diseases. *J Neural Transm Park Dis Dement Section*. 1990; 2:327-340.
19. Guan X, Xuan M, Gu Q, Huang P, Liu C, Wang N. *et al*. Regionally progressive accumulation of iron in Parkinson's disease as measured by quantitative susceptibility mapping. *NMR Biomed*, 2017, 30. doi:10.1002/nbm.3489.
  20. Daugherty A, Raz N. Age-related differences in iron content of subcortical nuclei observed in vivo: a meta-analysis. *Neuroimage*. 2013; 70:113-121.
  21. Fernandez B, Ferrer I, Gil F, Hilfiker S. Biomonitorization of iron accumulation in the substantia nigra from Lewy body disease patients. *Toxicol Rep*. 2017; 4:188-193.
  22. Foroutan P, Murray ME, Fujioka S, Schweitzer KJ, Dickson DW, Wszolek ZK. *et al*. Progressive supranuclear palsy: high-field-strength MR microscopy in the human substantia nigra and globus pallidus. *Radiology*. 2013; 266:280-288.
  23. Lee SH, Lyoo CH, Ahn SJ, Rinne JO, Lee MS. Brain regional iron contents in progressive supranuclear palsy. *Parkinsonism Rel Disord*. 2017; 45:28-32.
  24. De Reuck J, Capparos-Lebvre D, Deramecourt V, Defebvre L, Auger F, Durieux N. *et al*. Prevalence of small cerebral bleeds in patients with progressive supranuclear palsy: a neuropathological study with 7.0-Tesla magnetic resonance imaging correlates. *Folia Neuropathol*. 2014; 52:421-427.
  25. Tsunoda M, Yukino T, Koike H, Yoshino Y, Kamiyama M. Putaminal signal hypointensity on T2 weighted MR image in a patient with corticobasal degeneration. *Rinsho Skinkeigaku*. 2000; 40:59-61.
  26. Mizuno Y, Ozeki M, Iwata H, Takeuchi T, Ishihara R, Hashimoto N. *et al*. A case of clinically and neuropathologically atypical corticobasal degeneration with widespread iron deposition. *Acta Neuropathol*. 2002; 103:288-294.
  27. Savoiardo M. Differential diagnosis of Parkinson's disease and atypical parkinsonian disorders by magnetic resonance imaging. *Neurol Sci*. 2003; 24(Suppl 1):35-37.
  28. Matsusue E, Fujii S, Kanasaki Y, Sugihara S, Miyata H, Ohama E. *et al*. Putaminal lesion in multiple system atrophy: postmortem MR-pathological correlations. *Neuroradiology*. 2008; 50:559-567.
  29. Han Yh, Lee JH, Kang BM, Mun CW, Baik SK, Shin YI. *et al*. Topographic differences of brain iron deposition between progressive supranuclear palsy and parkinsonian variant of multiple system atrophy. *J Neurol Sci*. 2013; 325:29-35.
  30. Lee JH, Han YH, Kang BM, Mun CW, Baik SK, Shin YI. *et al*. Quantitative assessment of subcortical atrophy and iron content in progressive supranuclear palsy parkinsonian variant of multiple system atrophy. *J Neurol*. 2013; 260:2094-2101.
  31. Sugiyama A, Ito S, Suichi T, Sakurai T, Mukai H, Yokota H. *et al*. Putaminal hypointensity on T2\*-weighted MR imaging is the most practically useful sign in diagnosing multiple system atrophy: A preliminary study. *J Neurol Sci*. 2015; 349:174-178.
  32. Péran P, Barbagallo G, Nemmi F, Sierra M, Galitzky M, Traon AP. *et al*. MRI supervised and unsupervised classification of Parkinson's disease and multiple system atrophy. *Mov Disord*. 2018; 33:600-6008.
  33. Matsusue E, Fujii S, Kanasaki Y, Kaminou T, Ohama E, Ogawa T. *et al*. Cerebellar lesions in multiple system atrophy: postmortem MR imaging-pathologic correlations. *Am J Neuroradiol*. 2009; 30:1725-1730.
  34. Li X, Allen RP, Earley CJ, Cruz TE, Edden RAE, Barker PB. *et al*. Brain iron deficiency in idiopathic restless legs syndrome measured by quantitative magnetic susceptibility at 7 tesla. *Sleep Med*. 2016; 22:75-82.
  35. Frismand S, Salem H, Panouilleres M, Pélisson D, Jacobs S, Vighetto A. *et al*. MRI findings in AOA2: cerebellar atrophy and abnormal iron detection in dentate nucleus. *Neuroimage Clin*. 2013; 2:542-548.
  36. Ronsin S, Hannoun S, Thobois S, Petiot P, Vighetto A, Cotton F. *et al*. A new MRI marker of ataxia with oculomotor apraxia. *Eur J Radiol*. 2019; 110:187-192.
  37. Criscuolo C, Chessa L, Di Giandomenico S, Mancini P, Saccà F, Grieco GS. *et al*. Ataxia with oculomotor apraxia type 2: a clinical, pathologic, and genetic study. *Neurology*. 2006; 66:1207-1210.
  38. Anheim M, Monga B, Fleury M, Charles P, Barbot C, Salih M *et al*. Ataxia with oculomotor apraxia type 2: clinical, biological and genotype/phenotype correlation study of a cohort of 90 patients. *Brain*. 2009; 132:2688-2698.
  39. Salvatore E, Varrone A, Criscuolo C, Mancini P, Sansone V, Strisciuglio C. *et al*. Nigrostriatal involvement in ataxia with oculomotor apraxia type 1. *J Neurol*. 2008; 255:45-48.